

The Biomarkers in the Hyperbaric Oxygen Brain Injury Treatment Trial (BioHOBIT)

BIOSPECIMEN COLLECTION & PROCESSING

Overview

- 1. Specimen uniformity and quality
- 2. Site Equipment
- 3. Procedures
 - Kit Contents and Ordering
 - Sample Labelling
 - Sample Collection and Processing
 - Shipping Samples
 - Non-Conformance
- 4. Contact Information

Specimen Uniformity and Quality

GENERAL REMINDERS

Specimen Standardization and Quality

Most biomarkers are sensitive to *time* and *temperature*

- Standardization of processing across sites is key
- Specimens must be processed within 2 hours of collection
- Reference the BioSEND Biomarker Specimen Collection, Processing, and Shipment Manual as needed
- Do not replace or supplement any kit components without first receiving approval from BioSEND/NINDS

Questions? Email biosend@iu.edu

Site Consumables and Equipment

Sites will need to supply the following items:

- Gloves
- Alcohol wipes
- Butterfly needles
- Tourniquet
- Gauze pads
- Bandages
- Microcentrifuge tube rack
- Sharps bin and lid

- Crushed ice
- Pipettes and pipette tips
- Centrifuge capable of maintaining 4°C
- -80°C Freezer
- Dry ice

Procedures

MAINTAINING SPECIMEN UNIFORMITY AND QUALITY

Kit Contents and Ordering

- Kit and supply ordering will be through WebDCU
- Please ensure kits are marked as shipped in WebDCU, as this triggers the notification to BioSEND that your site needs more kits

Kit Contents and Ordering: Standard Kits

Standard Kit:

- Contains supplies for blood and CSF
- Can be used for any time point



Kit Contents and Ordering: Blood Kits

Shipping Kit:



Kit Contents and Ordering: CSF Kits

CSF: LP Tray:





Biospecimen Collection Protocol

Visit & Sample Type	Ohr/BL	8hr	24hr/D1	Post-HBO (one time between D2-D5)	D3	D5	D7	D14	D180
Plasma aliquots, 1500ul	4	4	4	4	4	4	4	4	4
Serum aliquots, 1500ul	4	4	4	4	4	4	4	4	4
Buffy Coat	1	1	1	1	1	1	1	1	1
CSF aliquots, 1500ul	4	4	4	4	4	4	0	0	0

Collection Volumes

Total blood, urine and CSF volumes

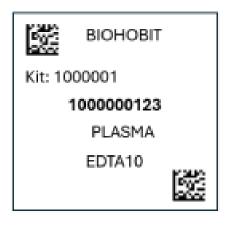
Sample Type	Amount
Whole Blood for Plasma and Buffy Coat	7 ml
Whole Blood for Serum	7 ml
Cerebrospinal Fluid	6 ml

Sample Labelling: Example Labels

Labels are provided by Indiana University

- Please check that all samples are properly labelled to ensure correct identification by IU
- If do not have enough labels to complete a visit, please contact IU immediately
- Labelling the tubes during processing prevents sample mix-ups





Sample Labelling: Label Placement

Please...

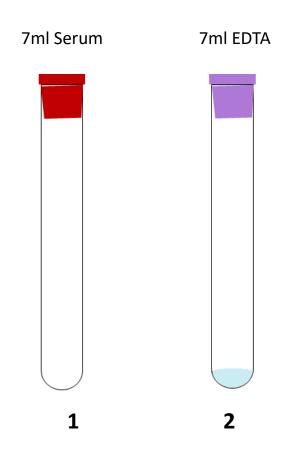
- Label all collection and aliquot tubes before cooling, collecting, processing, or freezing samples
- Label only 1 subject's tubes at a time to avoid mix-ups
- Wrap the label around the tube horizontally label position is important for all tube types
- Make sure the label is completely adhered by rolling between your fingers



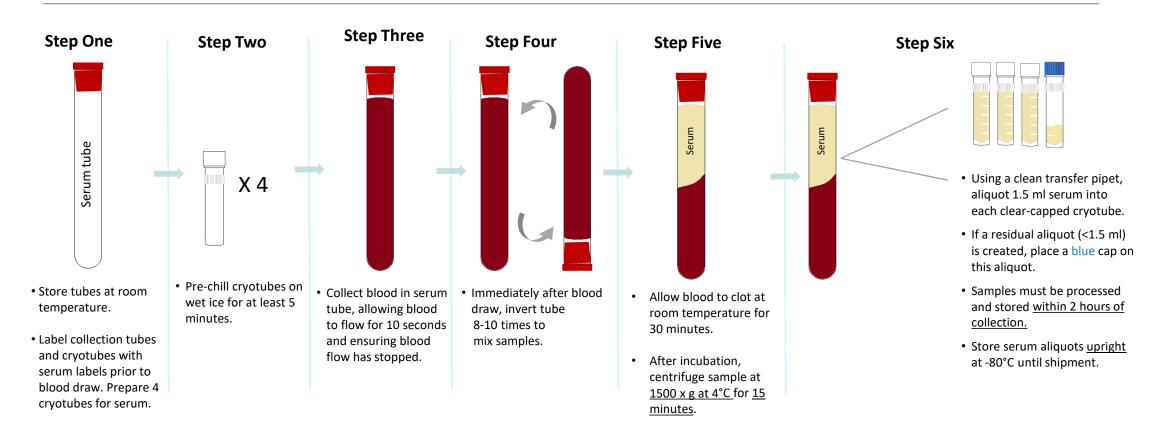


Sample Collection and Processing

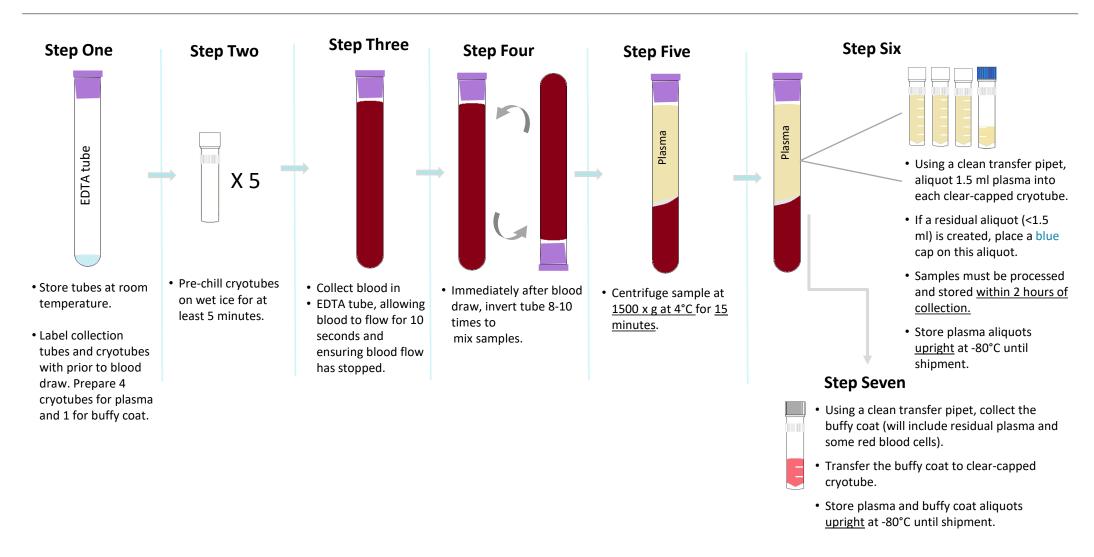
Blood Tube Draw Order



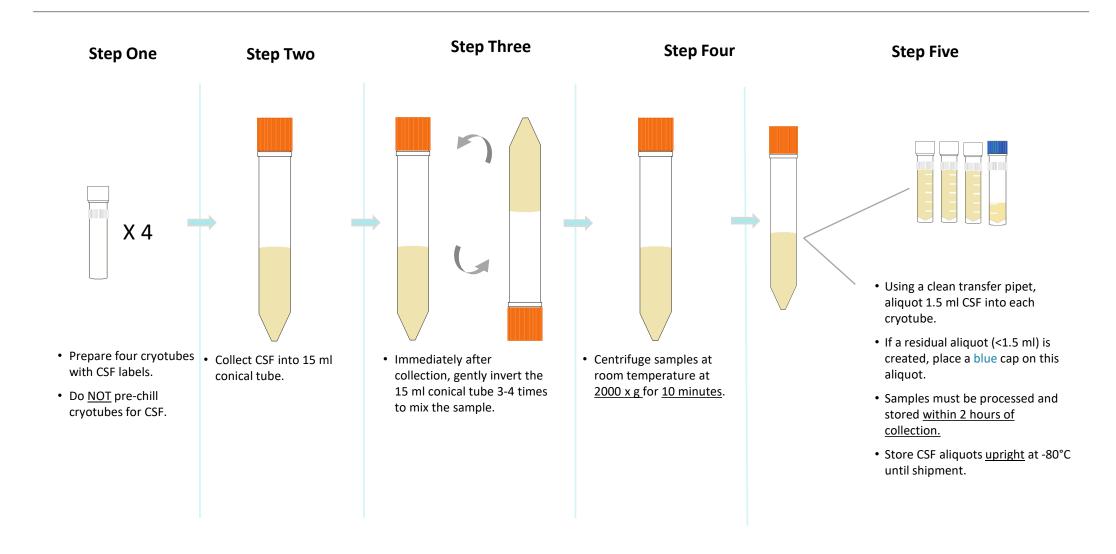
Sample Collection and Processing: Serum



Sample Collection and Processing: Plasma & Buffy Coat



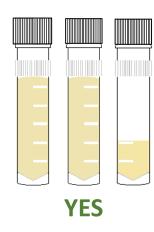
Sample Collection and Processing: CSF

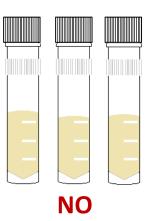


Sample Collection and Processing: Aliquots

Filling biomarker serum, plasma, and CSF aliquots:

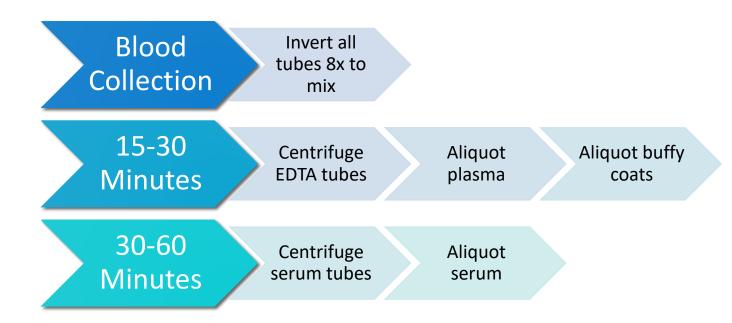
- Fill as many cryovials as possible to 1.5 ml (plasma, serum & CSF)
- Over-filled vials may burst in freezer!
- Ship ALL material to IU, even if final vial is less than standard volume





Sample Collection and Processing: Timeline

Timeline for blood processing



Sample Collection and Processing: Issue #1

Troubleshooting Blood Collection

Issue #1: Tube with little/no vacuum

- Always check expiration date on the tube before beginning blood draw and discard expired tubes
- Store tubes at "room temperature" extreme temperature can affect vacuum
- Keep extra vacutainer tubes from supplemental kit nearby during blood draw to replace "bad" tubes
- If this is a frequent occurrence, report tube type and lot number to IU.

Sample Collection and Processing: Issue #2

Troubleshooting Blood Collection

Issue #2: Hemolyzed serum and/or plasma caused by incorrect collection

Cause: Blood Collection Methods	Corrective Action				
Improper venipuncture site	Draw from median cubital, basalic, and cephalic veins from antecubital region of arm				
Prolonged tourniquet use	Tourniquet should be released after no more than 1 min, excessive fist clenching should be avoided				
Not allowing alcohol to dry on skin before venipuncture	Without touching, allow the venipuncture site to air dry				
Use of too large/small bore needle resulting in excess force applied to blood	Avoid using too small/large needle. Needle size dependent on the subject's physical characteristics & amount of blood to be drawn. Most commonly used sizes are 19 – 23.				
Pulling/pushing plunger too fast while drawing/transferring blood	Avoid drawing the syringe plunger too forcefully when collecting blood				
Ensure all blood collection assemblies are fitted securely, to avoid frothing					

For more information, visit: http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk_Jan2004_VS7167.pdf

Sample Collection and Processing: Issue #2 continued

Troubleshooting Blood Collection

Issue #2: Hemolyzed serum and/or plasma caused by incorrect processing

Cause: Sample Processing Methods	Corrective Actions				
Vigorous mixing/shaking	Gently invert blood collection tube when mixing additive with specimen, follow guidelines in Biologics Manual regarding number of times to invert each type of tube				
Not allowing serum to clot for recommended time	Serum tubes without clot activator should be allowed to clot for 60 min in a vertical position				
Exposure to excessive heat or cold	Keep samples at ambient temperature until processing				
Prolonged contact of serum/plasma with cells	Do not store uncentrifuged samples beyond recommended time				

For more information, visit: http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk_Jan2004_VS7167.pdf

Sample Collection and Processing: Sample Data

Specimen Collection and Processing Form

A copy of the Shipping Manifest should be included in every shipment to Indiana University







Ashely GARWOOD Sign Out

view: F181 Biosample Collection

Help

				JRF Fass	ed vennication	Create New Mic	onitor DCR Vie	w Audit Tra
CRF ID:	7755			Biosam	ple Collection		Rule Status:	DCR:
Site:	1047 University of Kentucky Hospital, Lexington, KY	Subject: 1229	Visit: Day	Submit:	15-Oct-2021 1 PETREY	4:29 Ronda	Accept:	Verify:
No.	Item Description				0	ata Value		
Q01	Biospecimen collection timepoint			ost injury				
Q02	Specimen collected			Yes				
Q03	Date of specimen collection							
Q04	Lab kit ID							
	Instruction: Blood tubes should be transported at room temperature. Allow no more than 90 minutes from time of blood draw to centrifuge start time.							
Q05	Blood sam	ple collected						
Q06		f blood draw						
	Serum Aliquots							
Q10	Time serum samples placed in freezer							
Q11	Aliquot S01 collected							
Q12	Aliquot S02 collected							
Q13	Aliquot S03 collected							
Q14	Aliquot S04 collected							
Q17	Reason any serum aliquot i							
	Plasma Aliquots							
Q20	Time plasma samples place							
Q21	Aliquot P01 collected							
Q22	Aliquot P02 collected							
Q23		03 collected						
Q24	Aliquot P04 collected							
Q27	Reason any plasma aliquot not collected							
	DNA (Buffy Coat)							
Q30	•	01 collected						
Q31	Time DNA place	ed in freezer						
	CSF aliquots Collect CSF sample as close to the blood collection	ae noeeihla						
Q40		SF collected	No o	Yes				
Q41		SF collected	0140	163				
Q42	Time CSF centrifu							
Q43	Time CSF centilio							
Q51		01 collected						
Q52	•	02 collected						
Q53		03 collected						
Q54	•	04 collected						
Q56	Reason any CSF aliquot i							
	Issues with processing or collecting samples		-					
Q57		ting samples	no staff a	/ailable a	t collection tim	ie		
GC						-		
Q57 GC	Any issues with processing or collect Genera	ting samples al comments			t collection tim	ie		

Last updated by Ronda PETREY on 15-Oct-2021 14:29

Sample Collection and Processing Form

Submission of the Shipping Manifest portion of the form serves as shipment notification to BioSEND

- Must be completed prior to shipment
- If samples are not marked as shipped in WebDCU, BioSEND will not receive notification that samples are shipped.
- Kit requests are triggered by WebDCU's tracking system of how many kits a site has remaining. If you do not ship kits to BioSEND, it will appear in the WebDCU system as if your site still has kits available. Regularly shipping samples to BioSEND ensures that your site continues to receive kit shipments.

Shipping Frozen Samples: Tips

Packing and Shipping Frozen Samples

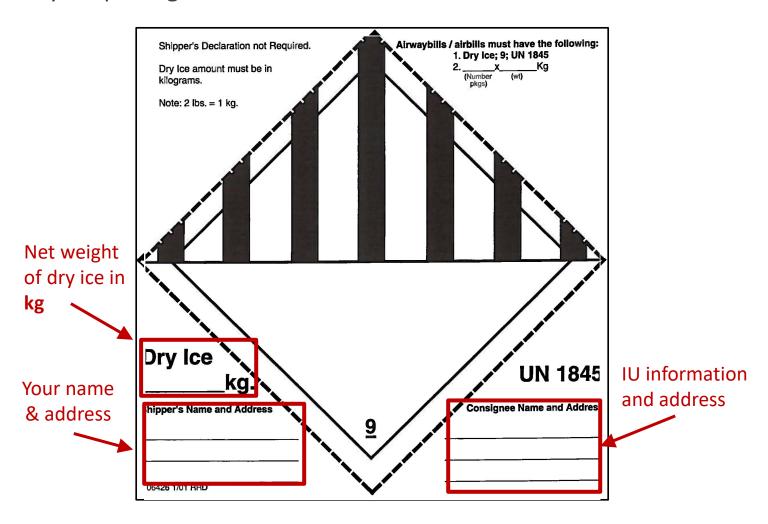
- Biomarker serum, plasma, buffy coats and CSF all ship frozen
- Ship frozen samples on dry ice
- Frozen samples should be shipped only Monday through Wednesday
- Always fill carton to top with dry ice
- Do not pack shipment until the day of pickup



Shipping Samples

Packing and Shipping Frozen Samples

Class 9 Dry Ice Label should not be covered with other stickers and must be completed, or UPS will reject/return your package!



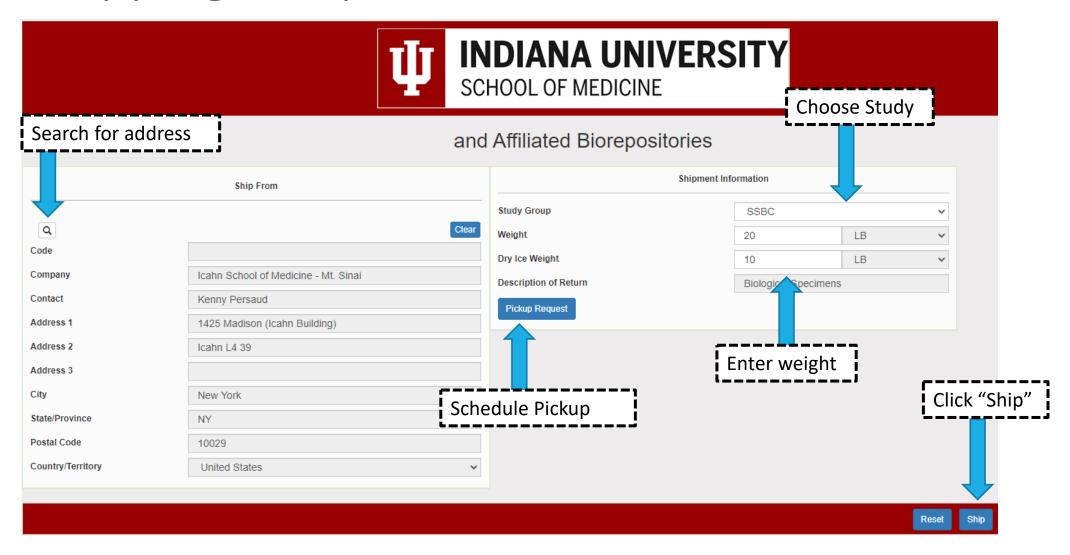
Shipping Samples: Frozen

Packing and Shipping Frozen Samples





Shipping Samples – UPS: https://kits.iu.edu/UPS



Shipping Samples via UPS

IU UPS ShipExec Shipping Portal

- Print out UPS air waybill
- Ensure all elements
 (barcode, return address, etc.) printed clearly
- Fold and insert UPS air waybill into clear plastic sleeve on package

JOHN SMITH INDIANA UNIVERSITY 410 WEST 10TH STREET INDIANAPOLIS IN 46202 2 LBS

1 OF 1

RS

SHIP TO:

SCHOOL OF MEDICINE 317-278-2694 INDIANA UNIVERSITY TK 217 351 W 10TH ST

INDIANAPOLIS IN 46202



IN 461 9-01



UPS NEXT DAY AIR

TRACKING #: 1Z 976 R8W 84 3985 8595

1



BILLING:

DESC: Biological Specimens RETURN SERVICE

Reference No.1: 4087277

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Shipping Samples: Closures

Date	Holiday				
January 1	New Year's Day				
3 rd Monday in January	Martin Luther King, Jr Day				
4 th Monday in May	Memorial Day				
June 19	Juneteenth (observed)				
July 4	Independence Day (observed)				
1 st Monday in September	Labor Day				
4 th Thursday in November	Thanksgiving				
4 th Friday in November	Friday after Thanksgiving				
December 25	Christmas				

Please also consider weather when shipping. UPS will post service updates on their webpage. Reach out to BioSEND if you an unsure if it is safe to ship.

Non-Conformance

Non-conformance to standard procedures may reduce the utility of the biospecimens:

- Not processing serum/plasma within 2 hours of collection allows for breakdown of certain proteins and small molecules
- Over/under centrifuging changes specimen composition



Non-Conformance Reporting con't

Most common non-conformance issues:

- Shipment notification not sent
- Samples shipped for weekend/holiday delivery
- Sample form incomplete/inaccurate
- Low volume
- Unlabeled or mislabeled tube(s)
- Sample hemolysis



Non-Conformance and Inventory Reporting

Most common non-conformance issues:

- BioSEND will notify sites directly of any issues upon receipt
- BioSEND will email sites a monthly inventory report of all samples received from that site to date
- If you are experiencing issues, please reach out to us for help! It is much easier to prevent an issue before sample collection & shipment than trying to fix it after the fact

Contacts

Indiana University

General Questions/Shipment Notifications:

biosend@iu.edu